Organic & Biomolecular Chemistry

Cite this: Org. Biomol. Chem., 2011, 9, 36

www.rsc.org/obc

PERSPECTIVE

Chiral separation by enantioselective liquid-liquid extraction

Boelo Schuur,^{†b,c} Bastiaan J. V. Verkuijl,^{†a} Adriaan J. Minnaard,^a Johannes G. de Vries,^{*a,d} Hero J. Heeres^{*b} and Ben L. Feringa^{*a}

Received 19th August 2010, Accepted 25th October 2010 DOI: 10.1039/c0ob00610f

The literature on enantioselective liquid–liquid extraction (ELLE) spans more than half a century of research. Nonetheless, a comprehensive overview has not appeared during the past few decades. Enantioselective liquid–liquid extraction is a technology of interest for a wide range of chemists and chemical engineers in the fields of fine chemicals, pharmaceuticals, agrochemicals, fragrances and foods. In this review the principles and advances of resolution through enantioselective liquid–liquid extraction including host–guest chemistry, extraction and phase transfer mechanisms, and multistage liquid–liquid extraction processing. Then the literature on enantioselective liquid–liquid extractant classes are considered: crown ether based extractants, metal complexes and metalloids, extractants based on tartrates, and a final section with all other types of chiral extractants.

Introduction

The availability of enantiopure compounds is of prime importance for the pharmaceutical¹ and also to some extent for the agrochemical, flavour and fragrances industries.² They may be obtained either *via* a synthetic approach,³ or *via* the separation of racemates (Fig. 1).^{4,5} The synthetic routes can be based on natural compounds, fermentation or asymmetric synthesis.

The chiral pool strategy uses chiral compounds from nature or products derived thereof (*e.g.* from fermentation processes⁶). Examples of industrial fermentations are the production of various acids (*e.g.* lactic acid, citric acid),⁷ penicillin,⁸ and α -amino acids.⁹ Usually the production using this strategy is relatively cheap. Unfortunately, not all enantiopure precursors can be obtained from natural sources. When the chiral pool is not an option, conversion of prochiral compounds into chiral compounds by asymmetric synthesis¹⁰ is an attractive alternative. Although asymmetric catalysis is intrinsically very powerful, its use in practice may be hampered by high catalyst costs and limited development time caused by time-to-market pressure.^{11,12}

^cEindhoven University of Technology, Chemical Engineering and Chemistry Department, P.O. Box 513, 5600 MB, Eindhoven, The Netherlands. E-mail: B.Schuur@tue.nl; Fax: +31 40 246 3966; Tel: +31 40 247 4716

^dDSM Innovative Synthesis BV – A unit of DSM Pharma Chemicals, P.O. Box 16, 6160 MD, Geleen, The Netherlands. E-mail: Hans-JG. Vriesde@DSM.com; Fax: +3146 476 7604; Tel: +31 46 476 1572

[†] These two authors contributed equally to this manuscript.



Fig. 1 Available methods for acquiring enantiopure compounds.

Enantiopure compounds may also be obtained by racemic synthesis followed by enantiomer separation. The time-to-market for this strategy is potentially shorter than in asymmetric catalysis, provided the separation of enantiomers can be done with a technique that is broadly applicable to structurally diverse substrates. A facile synthesis of racemic compounds followed by a racemate separation using a broadly applicable technology that is easily developed is possibly very beneficial. Separation of racemates on industrial scale is usually based on resolution by crystallization.¹³⁻¹⁷ Although resolution by crystallization is the most frequently used route to enantiopure compounds, the main drawbacks are the low versatility and excessive solids handling and a maximum yield of 50%,¹⁸ unless racemization of the undesired enantiomer can be applied.¹⁹

Numerous laboratory techniques are available for enantioseparation.²⁰⁻²⁴ Typically, for scaling up of successful and very broadly applicable laboratory techniques such as chromatography²⁵ and capillary electrophoresis,²⁶ high capital

^aUniversity of Groningen, Stratingh Institute for Chemisty, Nijenborgh 4, 9747 AG, Groningen, The Netherlands. E-mail: B.L.Feringa@rug.nl; Fax: +31 50 363 4278; Tel: +31 50 363 4296

^bUniversity of Groningen, Chemical Engineering Department, Nijenborgh 4, 9747 AG, Groningen, The Netherlands. E-mail: H.J.Heeres@rug.nl; Fax: +31 50 363 4479; Tel: +31 50 363 4484

investments are required.²⁷ Nevertheless, it has been argued that for small volumes of single enantiomers needed in early development stages, the high separation costs by chromatographic techniques are still small compared to the total drug development costs.²⁸ Some impressive preparative chiral separations have been demonstrated using centrifugal partition chromatography,²⁹ and simulated moving bed chromatography.^{30,31} Another strategy for chiral separations is to use membrane-based approaches.^{12,32-35} When applying immobilized selectors in (liquid) membranes, the amount of selector needed can be reduced greatly. Limitations of this technology are the relatively low transport rates through the membranes requiring high membrane areas, delicate pressure control, and the risk of fouling.

In liquid-liquid extraction, diffusion and convection are the transport mechanisms, and high transport rates should be achievable.³⁶ Liquid-liquid extraction is a mature technology that can easily be operated in a continuous countercurrent mode to fractionate the racemate into its enantiomers,³⁷ which is advantageous when scaling-up. This possibility to operate at all scales, from laboratory separations to bulk processes in the chemical industry, makes the use of liquid-liquid extraction for enantioseparation especially interesting. The use of liquid-liquid extraction for enantioseparation has been known since 1959,³⁸ and the first articles in the English literature appeared in the late 1960's.³⁹⁻⁴² Enantioselective liquid-liquid extraction (ELLE) is an attractive alternative for chromatography and resolution by crystallization that, based on the scientific literature, still needs development towards commercialization. Studies cited in the literature deal mainly with the introduction of novel extraction systems. Only a few engineering studies have been reported.^{36,43-47} In the past 15 years, two reviews on aspects of chiral resolution by extraction have appeared, the first one (1994) with a strong emphasis on extraction chemistry,48 and the second one (2001) on extraction technology.³⁷ In the current overview, the reader is first introduced to the principles of ELLE and the development strategy for successful application of ELLE in (continuous) industrial processes. Ample literature is available on liquid-liquid extraction equipment.⁴⁹ However, the equipment that has typically been used by researchers in the field is different from traditionally and industrially applied equipment. Therefore, a concise overview of extraction equipment is provided. Next, an overview of ELLE chemistry, categorized according to chiral extractant classes, is presented as well as an overview of hosts. Finally, conclusions and an outlook on the field are given.

Principles and application of enantioselective liquid–liquid extraction

ELLE is closely affiliated to the large field of host–guest chemistry⁵⁰ and chiral recognition can be regarded as a mere practical application of the developments in this wide field. Other closely related applications are the optical resolution of racemic compounds by inclusion complexation.^{51–54} and resolution by distillation with inclusion compounds.⁵⁵ ELLE combines the concepts of enantiomeric recognition⁵⁶ and solvent extraction^{57–59} in a single technique. In this section, the principles underlying ELLE and the approaches for application of ELLE in multistage extraction processes will be introduced.

Host-guest chemistry and chiral recognition

The principle of enantiomeric recognition is essential for enantioselective complexation and thus for ELLE. Without enantiomeric recognition, enantioselective processes are not possible. Encounter complexes of the extractant with the enantiomers are formed as a result of intermolecular interactions.⁶⁰ These intermolecular interactions may include ion pairing, hydrogen bonding, π - π interactions, dipole and Van der Waals interactions.⁵⁶ The complexation of a chiral host with a chiral guest bearing one stereocenter can be schematically represented as the guest binding with groups B, C and D to the groups B', C' and D' of the host (Fig. 2, left). The opposite enantiomer will bind to two out of three sites (Fig. 2, right). The three point attachment (TPA) model states that the complex in Fig. 2 (left) is the preferred one.⁶¹ If the interactions, which are presented as dotted lines, represent not just attracting interactions, but also repulsive interactions, the TPA model is modified to the three point interaction (TPI) model.^{62,63}



Fig. 2 The three point model for the complexation of both substrate enantiomers to a chiral host.

The field of enantiomeric recognition was boosted in the 1970's by Lehn and Cram^{50,64} and has led to a broad spectrum of host–guest chemistry. A wide variety of discriminative interactions has been reported. The field has been extensively reviewed and various approaches have been discussed: host–guest chemistry in general,⁶⁵ anion recognition,^{66–68} peptide and protein recognition,⁶⁹ carbohydrate recognition⁷⁰ and recognition of carboxylic acids,⁷¹ neutral molecules⁷² and amino compounds.⁷³

Extraction and phase transfer mechanisms

In addition to the chiral recognition principle, it is also essential for ELLE that there should be two (at least partially) immiscible phases, normally an aqueous and an organic phase. A typical ELLE system initially has the substrate predominantly confined in the aqueous phase (Fig. 3). After adding a lipophilic host or extractant, which prefers to be confined in the organic phase, a host-mediated phase transfer of the substrate occurs. In the example illustrated in Fig. 3, interaction of the host with the (S)-enantiomer is favored over interaction with the (R)-enantiomer. As a result, the organic phase will be enriched with the (S)-enantiomer and the aqueous phase enriched with the (R)-enantiomer.

Without going into mechanistic detail, the two main mechanisms given in the extraction literature⁴⁶ are introduced here briefly. The two different types of extraction mechanism that have been proposed for ELLE are the homogeneous ligand addition mechanism, involving a homogeneous reaction in either one of the liquid phases between host and guest, and the interfacial ligand exchange mechanism.^{46,75} The two mechanisms are depicted in Fig. 4.



Fig. 3 Extraction and enantiomeric recognition in a biphasic system. Symbols: (*S*)-enantiomer, (*R*)-enantiomer, (*S*): host. Taken from ref. 74 with permission, © 2009, ACS.



Fig. 4 Interfacial ligand exchange mechanism of solute A with extractant C (left) and homogeneous organic phase ligand addition mechanism (right).

The main difference between the two models is the locus of the complexation reaction. The interfacial reaction model obviously applies when the reaction takes place at the interface. This situation is likely to be valid when the guest is insoluble in the organic phase and the host is insoluble in the water phase. This type of mechanism is frequently observed in the field of metal-ion extractions,76,77 and is characteristic for ligand exchange extraction systems. Ligand exchange systems may also be appropriate for ELLE of ionic guests. When either the host or guest is soluble in the other phase, homogeneous reactions are also possible. The homogeneous reaction model is appropriate for ELLE systems with a substrate addition mechanism, and the solutes are at least slightly soluble in the extract phase (which is usually the case, as the solutes are typically organic compounds). This type of mechanism has been reported, for instance, for extractions of organic acids from aqueous fermentation broths.78,79

Understanding of the complexation mechanism is important when aiming at a process optimization. Typically, the substrates (α -amino acids, amino alcohols, and carboxylic acids) can exist in a neutral form, and in a (de)protonated form, depending on the actual pH in the aqueous phase. When the extraction proceeds through an interfacial anion exchange mechanism, the highest distributions are found at a pH where the guests prevail in the deprotonated form. When instead a homogeneous organic phase ligand addition mechanism is operative, the highest distributions may be found at a pH where the neutral form of the guest prevails.

Approaches for ELLE application in multistage processes

A key element in developing the technology of ELLE from the principles discussed above into a satisfactory separation process is the multistage countercurrent process approach. With this approach, it is not necessary to achieve a complete separation of enantiomers in a single stage and therefore also the selectivity requirements are not as extreme as in. for example, asymmetric catalysis. As a result, it is more likely that a single extractant can be applied to separate a broader range of racemates, which is desired.27 With versatile extractants for racemate separations, the development times for new synthetic routes of chiral compounds can be shortened. Thus, with the multistage approach, highly enantiopure compounds are obtainable in high yields using selectors that display only moderate selectivity. Several configurations may be applied, such as cocurrent multistage extraction,⁸⁰ countercurrent multistage extraction,⁸¹ or countercurrent multistage fractional extraction,⁴⁷ which may be considered as the most efficient configuration. Fig. 5 displays a scheme of a fractional extraction setup with a back-extraction section to recover the host to enable recycling. In fractional extraction, the feed enters the cascade of extractors or the extraction column at an intermediate stage. The stages from the feed stage F to stage N are called the strip section, the stages 1 to F form the wash section. This configuration allows efficient extraction of the desired enantiomer in the strip section, whereas the co-extracted enantiomer that is not desired is washed out of the extract stream in the wash section. The extract stream with the desired enantiomer leaves the wash section from stage 1, and enters the back-extraction section where the host is recovered to re-enter the strip section in stage N. The recovered enantiomer leaves the process in the back-extraction stream. If the enantiopurity in this stream is very high, part of the stream may be refluxed to join the wash stream and re-enter in stage 1 of the wash section. As a consequence, the increased concentration of the preferred enantiomer in the aqueous stream at the start of the cascade prevents the preferred enantiomer already complexed to the host in the organic phase from back-extracting into the aqueous phase. In this way the enantiomeric excess increases, so that a reduction of the number of extraction stages, and of the required amount of solvent is possible.



Fig. 5 Multistage fractional extraction scheme with back-extraction section to recover the host.

Several stages may be identified in the development of a multistage fractional extraction process. The first and probably the most essential step is the selection of a good host. Host selection is not easy and although over the years numerous hosts have been identified for the extraction of several substrate classes such as α -amino acids, amino alcohols and carboxylic acids, most hosts show only reasonable selectivity for a limited number of racemates. Besides being selective, the host–substrate complexation needs to be reversible and the host needs to be confined in a single phase as much as possible. These properties are vital for the application in a fractional extraction cascade. The hosts that have been reported thus far are discussed further on.

After a suitable host has been selected, single stage batch experiments are performed with the aim of the elucidation of the extraction mechanism and single stage extraction modeling. With an appropriate single stage model, the distribution of the enantiomers may be determined as function of the process conditions. Also, the single stage model parameters may be used in modeling multistage processes in order to determine the minimum wash, extract, and back-extraction flows to separate the enantiomers in the feed stream, and to determine the minimum number of stages for this separation. Heeres et al. demonstrated that ELLE is possible in continuous centrifugal contactor separator equipment.⁸² These continuous centrifugal contactor separators (CCS) seem highly suitable for ELLE, because the liquid hold-up is limited, and only a small amount of the precious host needs to be available in order to separate larger amounts of enantiomers in a continuous flow mode. Batch experimentation techniques were applied to develop an equilibrium stage model for the ELLE of racemic 3.5-dinitrobenzovl-leucine using a *Cinchona* alkaloid type of host.75 The applicability of equilibrium stage modeling in this continuous process was also demonstrated. A single extraction stage corresponded to an equilibrium stage.

The relationship between the minimal number of required fractional extraction steps (N_{\min}) for full separation of both enantiomers of the substrate and the desired ee (or fraction equivalents, defined by x_R and x_S) of the substrate and the operational selectivity (α_{op}) of a single extraction, is given by the Fenske equation (eqn (1)).

$$N_{\min} = \frac{\ln\left(\frac{x_{R}/(1-x_{R})}{x_{S}/(1-x_{S})}\right)}{\ln\alpha_{op}}$$
(1)

The relationship between $N_{\rm min}$ and $\alpha_{\rm op}$ is depicted in Fig. 6 (left). At an $\alpha_{\rm op}$ of 1.5, $N_{\rm min}$ is still 25. $N_{\rm min}$ drops exponentially as $\alpha_{\rm op}$ increases. This graph illustrates that full separation is already possible at moderate levels of $\alpha_{\rm op}$ by using a fractional extraction CCS cascade with a reasonable number of fractional steps. Furthermore, it is evident that increasing $\alpha_{\rm op}$ beyond 7 provides only a slight decrease in $N_{\rm min}$. This illustrates the strength



Fig. 6 The minimal number of fractional extraction stages (N_{\min}) required for ee = 99% as a function of α_{op} (left) and the minimal number of stages required for a system with $\alpha_{op} = 7.0$ as a function of ee (right).

of the technology, as it is evident that with a limited number of stages a racemate can be separated, provided that there is a modest selectivity.

In Fig. 6 (right), the relationship between the desired ee and $N_{\rm min}$ is depicted for an $\alpha_{\rm op} = 7.0$. It is clear that only 4 stages are required to obtain 95% ee, and with 7 stages an ee as high as 99.8% ee is obtained.

The above mentioned examples of continuous extraction modeling with equilibrium stage models both involve discrete stages (like those displayed in Fig. 5). In more traditional extraction columns, no discrete stages can be identified, but a certain height corresponds with one theoretical equilibrium stage. For extraction columns, the number of stages calculated with an equilibrium model can thus be translated into the required height of the column using the height equivalent of a theoretical stage (HETS)⁴⁹ approach. The HETS should be determined experimentally.

Single stage equilibria

The most common way to describe the performance of an ELLE process is using the ee. However, the ee alone does not fully describe the performance. This subsection lists the definitions required for adequate performance determination.²⁷ The ee is a typical example of an operational definition, and not an intrinsic one. Operational definitions describe experimental observations. Another example of an operational definition is the operational selectivity of an ELLE system. The operational selectivity is defined as the ratio of the distributions of the enantiomers, when the (R) enantiomer is preferentially extracted:

$$\alpha_{\rm op} = \frac{D_R}{D_S} \tag{2}$$

In eqn (2), the distributions are defined as:

$$D_{i} = \frac{[i]_{\text{org, all forms}}}{[i]_{\text{aq, all forms}}}; \qquad (i = R, S)$$
(3)

Both the aqueous phase concentration of the enantiomers and the organic phase concentration is the sum of the concentrations of all forms present in that phase, e.g. for the aqueous phase concentration of a weak Brønsted acid both the acid and its conjugated base should be taken into account. For example, the N-protected dinitrobenzoyl α -amino acids behave as weak acids, and depending on the pH either the neutral form or the deprotonated form is the predominant species.75 Similarly, in case of the homogeneous complexation model, the total concentration of the enantiomers in the organic phase is the sum of the host-guest complex concentrations and the concentrations of the free enantiomers in the organic phase (the interfacial ligand exchange model is most likely with almost zero solubility of the free enantiomers in the organic phase, hence only host-guest complexes are observed in the organic phase and no free enantiomers). The operational definitions are important to determine the actual performance of the extractive system and to establish the product purity, but do not give information on the intrinsic enantioselectivity of the extractant. The intrinsic selectivity of an ELLE system is defined as the ratio of the equilibrium constants, e.g. for preferential complexation of the host with the (R) enantiomer:

$$\alpha_{\rm int} = \frac{K_R}{K_S} \tag{4}$$

the exact definitions of the complexation equilibria K_i depend on the mechanism of extraction (Fig. 4) and may be defined either for a heterogeneous system when the interfacial ligand exchange mechanism applies, or for a homogeneous system when the homogeneous ligand addition mechanism applies. In the case of homogeneous complexation, the concentration of the free enantiomers is related to the free enantiomer concentration in the aqueous phase (ionic species hardly partition):

$$[R]_{\rm org} = m[R]_{\rm aq} \tag{5}$$

Thus, using the aqueous phase dissociation/protonation relations, the partition coefficient and the complexation equilibrium relations, a single stage can be described fully intrinsically. An example of such an intrinsically defined single stage extraction is given in Fig. 7.



Fig. 7 Homogeneous ligand addition mechanism as described by Heeres *et al.*⁷⁵ \bigcirc 2008, ACS.

Enantioselective liquid-liquid extraction equipment

Traditional equipment for liquid-liquid extraction used in the chemical industry covers a wide range, both in types and in sizes.⁴⁹ Selection of the most suitable type is dependent on the process conditions and constraints and may be supported by selection charts.⁸³ There are two basic types of extractors, the mixer-settlers that form discrete stages and are used for the bulkiest of all separation processes, and the extraction columns that are most broadly applied in all kinds of extraction processes. Extraction columns are considered as differential equipment, as no discrete stages can be distinguished. Therefore, the number of stages in a column is determined by expressing the height equivalent of a theoretical stage (HETS). Many types of columns are available with a broad range of internals used to enhance the mass transport facilitating the separation.⁵⁷ For special needs, such as short contact times to avoid product degradation, numerous types of centrifugal extractors have been developed,⁸⁴ some resembling the discrete stages in traditional mixer-settlers (e.g. the centrifugal contactor separators) and others resembling the differential contacting equipment (e.g. the Podbielniak centrifugal extractor).85

Bulk liquid membranes

Industrial equipment is less suitable for fundamental research on extraction mechanisms. Therefore, researchers have come up with innovative types of equipment that allow for demonstration of enantioselectivity or elucidation of intrinsic parameters. Fundamental research on extraction kinetics is commonly performed using Lewis type of stirred cells.^{76,86} Next to the traditional Lewis cell, novel devices have been developed for ELLE characterization. The first novel device that was especially designed for ELLE purposes was the chiral resolution machine developed by Cram and coworkers⁸⁷ (Fig. 8).



Fig. 8 Cram's catalytic resolving device, reproduced from ref. 87 with permission. \bigcirc 1979, ACS.

This simple resolution device is fed with aqueous racemate in the middle compartment, and the two U-tubes filled with heavier chlorinated organic liquids that contain each one of the enantiomers of the host. The individual enantiomers from the racemate travel preferentially through one of the chiral host phases, depending on the enantiomer of the host applied and end up in the outer aqueous receiving compartments.

The groups of de Mendoza⁸⁸ and De Vries⁸⁹ have applied single U-tubes with only one enantiomer of the hosts to examine the resolving process. With the U-tube type of resolving equipment, having exactly determined interfacial areas, mass transfer characteristics could be studied, however, they are mostly applied without a stirrer in the feed and receive phase. The use of a device as in Fig. 8 could ensure well mixed aqueous phases in order to determine mass transfer characteristics properly.

Supported liquid membranes

Abe *et al.* reported the continuous separation of mandelic acid enantiomers with their liquid particle extractor, which may be considered as a continuous chromatographic device.⁹⁰ Although the step from batch to continuous separation is worth mentioning, the device has never matured to preparative scale, most likely due to the large solvent flows required. More recently, supported liquid membrane technology (see Fig. 9) was reported as another type of continuous enantioseparation process.³⁴ With this technology very high purities may be obtained with only limited amounts of precious hosts; however the practical use is limited by the risk of fouling, loss of extract phase, and the slow transport rates.

Centrifugal contactor separators (CCS)

Until recently, the state of the art in ELLE was limited to a demonstration of the proof of principle for ELLE using the



Fig. 9 Supported liquid membrane flow.

above discussed types of equipment for batch-wise operation, or applying chromatographic or membrane approaches. The research on continuous ELLE processes in liquid–liquid biphasic systems to design multistage countercurrent fractional extraction processes for separation of racemates into single enantiomers with only moderate extractant selectivity had not been reported. This is possibly due to the large volumes required in the traditional types of continuous extraction equipment. Making use of centrifugal contactor separators (CCS), only small amounts of the precious chiral extractants are required for multistage continuous separation of enantiomers by extraction, as demonstrated by Heeres *et al.*^{91,92}

In this CCS (Fig. 10), the organic and aqueous liquids enter the device separately in the annular mixing zone, where they move down by means of gravity and become intensely mixed due to the high shear stress between the fast rotating centrifuge and the static wall of the device, allowing interfacial mass transfer and enantioselective partitioning when a chiral, enantioselective extractant is used. It was demonstrated that because of the



Fig. 10 Centrifugal contactor separator, courtesy of CINC Solutions, Doetinchem, The Netherlands.

intensive mixing and the fast reaction, one experimental stage corresponded with an equilibrium stage. Next, the dispersed liquids are moved to the centrifuge through a hole in the bottom. The liquids are efficiently separated by the large centrifugal forces. The device is thus a centrifugal version of a mixer–settler with the advantage that through use of centrifugal force only small amounts of liquids are needed.

Extractant classes

Crown ether based reactive extractants

The crown ether based systems are among the most enantioselective reactive extractants known. The discovery, synthesis and application of crown ethers were pioneered by Pedersen.^{93,94} The crown ethers were provided with chiral moieties and applied as chiral hosts by the groups of Lehn⁹⁵ and Cram.⁹⁶ The versatility of crown ether based extractants in ELLE may be limited to ammonium salts, but their operational selectivities up to 31 make it the most important extractants in the field.

Cram's BINOL crown ether system

Cram's dilocular host contains a macrocyclic uncharged crown ether carrier sided by two BINOL moieties of which one BI-NOL has two methyl directing side-groups at the 3,3'-positions (Fig. 11).⁹⁶ It gives selectivities up to 12 for amine salts,⁸⁷ and up to 31 for α -amino acid ester salts.⁹⁷



Fig. 11 Cram's dilocular crown ether host.

The enantioselective recognition is restricted to primary amine salts, since the ammonium cation complexes in a tripodal configuration to the crown ether. Steric hindrance by side groups connected to the 3,3'-positions of one BINOL backbone direct the bulky groups into a favorable position. When the D-enantiomer of an α -amino acid ester complexes with a (R,R)-host, the R substituents in the α -amino acid ester occupy the most spacious cavity, and the NCH hydrogen rests against the chiral barrier of the host formed by the directing methyl group of the host (Fig. 12).



Fig. 12 The complexation of an α -amino ester salt with the dilocular crown ether host. Schematic bars represent binaphthyl moieties.

In subsequent years, Cram *et al.* managed to design a chiral host with only one chiral element (Fig. 13).⁹⁸ The directing



Fig. 13 Chiral crown ether based host with only one chiral element, A = directing side group.

side-groups (A) turned out to be vital for chiral recognition, giving low selectivity for A = H. The highest selectivity was observed for A = Ph, ranging from 3.9 up to 19.5 for the ammonium salts of α -amino acids and α -amino acid methyl esters as substrates. The selectivity towards phenylglycine was increased to 23.4 using a mixture of acetonitrile and chloroform as the organic phase.

Guanidinium-crown ether tethered systems

The guanidinium-crown ether tethered system published by de Mendoza *et al.*⁹⁹ is a beautiful example of the use of multiple functionalities to achieve chiral recognition of α -amino acids (Fig. 14, top). The molecule was designed with four possible interaction sites, of which one proved not to be essential. One important issue was that the functional groups interacting with the ammonium and carboxylate, respectively, need to be non-self-complementary. Therefore, a crown ether and a guanidinium function were used to prevent the host from internal collapse. The naphthalene ring was introduced for extra π - π interaction with the side chains of aromatic α -amino acids. Finally, the structure was made homochiral to achieve enantioselective recognition.



Fig. 14 De Mendoza's guanidinium-crown ether tethered host (top), and proposed complexation of the host complexed with L-tryptophan (bottom).

The structure proposed in 1992 shows a 1:1 complex between the (*S*,*S*) structure of the host (depicted in Fig. 14, bottom) and L-Trp. However, structural and molecular dynamics studies on the same complex showed that in the case of L-tryptophan, the indole moiety is folded over the aza-crown ether (Fig. 14, bottom), thereby excluding some additional polar surface from interactions with the apolar solvent (chloroform).⁸⁸ In the case of D-tryptophan the indolyl ring dangles into the solvent.

Lipophilic crown ethers

The ELLE of amine picrates using lipophilic crown ethers (Fig. 15, upper panel) has been reported by Costero *et al.*¹⁰⁰ and Nazarenko *et al.*¹⁰¹ with selectivities up to 3. Picrates (Fig. 15, lower panel) are salts or esters of picric acid and known to be soluble in both the aqueous and organic phase.¹⁰²



Fig. 15 Crown ethers (upper panel) for extraction of picrates (lower panel).

Azophenolic crown ethers

An important member among the family of chiral crown ether hosts is the azophenolic crown ether shown in Fig. 16 (left). Azophenolic crown ethers were synthesized by Naemura *et al.*¹⁰³ to study complexation in homogeneous solutions of chiral amines and amino alcohols.¹⁰⁴ The crown ether is able to form a tripod-like complex with a neutral amine (Fig. 16 (right). The acidic proton of the phenol forms a hydrogen bond with the amine. The acidity of the phenol is increased by the electron withdrawing nitro groups. This increased acidity is presumed to enhance the complexation of the azophenolic crown ether with a primary amine.



Fig. 16 Azophenolic crown ether host (left) and the proposed complexation structure of an azophenolic crown ether with a neutral primary amine (right).

This system has been extensively studied by De Haan *et al.*⁴⁵ With phenylglycinol, an intrinsic selectivity of up to 12 was achieved, although other amines show lower selectivity (1.5–3.2).

Metal complexes and metalloids as reactive extractants

Cu(II)proline derivative complexes. In 1979, the resolution of α -amino acids using ligand exchange chromatography with a chiral mobile phase was reported by Hare and Gil-Av.¹⁰⁵ Racemic α -amino acids were separated with the addition of a copper(II) proline complex to the eluent using a cation-exchange column. Gübitz *et al.* described the synthesis of a chiral stationary phase (Fig. 17, left) with the copper(II)proline complex.¹⁰⁶ α -Amino acids can complex to the copper(II) cation in a bidentate fashion. Separation of α -amino acid racemates was achieved by the virtue of different complex stabilities between the mixed complexes with the D- (Fig. 17, right) and L-form, respectively.¹⁰⁷ The hydroxyl function in the side chain proved to be important for separation, as absence of the hydroxyl function significantly lowered the selectivity. A range of α -amino acids could be separated with this method.¹⁰⁸



Fig. 17 Proposed complex of the chiral stationary phase (left) and the diastereomeric complex between copper(II)(L-proline) as the stationary phase and D-phenylalanine (right).

Takeuchi *et al.*¹⁰⁹ were the first to use this system for enantioselective solvent extraction of neutral α -amino acids. *N*-Alkylated-L-proline derivatives were used as ligands for copper, to keep the host confined into the organic phase. The use of L-4hydroxyproline gave an increase in operational selectivity (Fig. 18). The complexing constants of the α -amino acid enantiomers were determined by variation of the cupric-ion concentration with respect to the pure enantiomers. The resulting intrinsic selectivities ranged from 1.8 to 4.5 for non-aromatic α -amino acids.



Fig. 18 Copper(II)(N-alkyl-4-hydroxyproline)₂.

Pickering and Chaudhuri^{44,110} demonstrated the use of two different extractants Cu(II)(*N*-dodecyl-(L)-hydroxyproline)₂, Cu(II)(C-12Hyp)₂ and (*S*)-bis(phenylnaphtho)-20-crown-6) for the ELLE of phenylalanine. The selectivity using the Cu(II)(C-12Hyp)₂ complex was in the range reported earlier by Takeuchi *et al.*,¹⁰⁹ establishing the versatility of the system. De Haan *et al.* studied the Cu(II)(*N*-(2-hydroxydodecyl)-L-hydroxyproline) complex as an extractant for a series of primary amines.²⁷ They found dependency between distribution and operational selectivity as well as the pH. Similar results of these correlations were also shown by Pickering¹¹¹ and Ding.¹¹² The Cu(II)(*N*-(2-hydroxydodecyl)-L-hydroxyproline) complex has a selectivity of 1.8 for norephedrine while for other amines an α_{op} below 1.2 was obtained. Dimitrova and Bart have demonstrated the application of microemulsions for the enantioselective extraction of phenylalanine and tryptophan with copper hydroxyprolines.¹¹³

Lanthanide(III)tris(\beta-diketonate) complexes. Tsukube *et al.* reported the use of chiral lanthanide(III) tris(β -diketonates) in the ELLE of unprotected amino acids.¹¹⁴ Some chiral lanthanide(III) tris(β -diketonates) are known to act as effective receptors for anionic substrates in solutions and are frequently used as chiral shift reagents in NMR spectroscopy.¹¹⁵ The negatively charged and highly coordinated complexes can interact and bind the ammonium part of the α -amino acid in an intramolecular fashion *via* electrostatic interaction (Fig. 19).¹¹⁶



Fig. 19 The proposed complexation of a zwitterionic amino acid with a lanthanide tris(β -diketonate) (Q represents β -diketonate ligand).

Upon variation of the lanthanide cation, opposing trends were observed for distribution and enantioselectivity.¹¹⁷ The correlation between the ion size of the central metal cation (Pr(III) > Eu(III), Eu(II) > Yb(III)) and the extractability was negative, whereas the enantioselectivity increased. This may be due to the smaller cation providing closer contact with the α -amino acid to enhance the α_{op} up to 2.2 in the case of the ytterbium complex **2e** (Fig. 20) with a (+)-camphor-derived ligand for the extraction of tryptophan.¹¹⁸



Fig. 20 Lanthanide complexes with (+)-camphor derived ligand.

Metalloporphyrins. Inoue *et al.* designed a host based on a metalloporphyrin,¹¹⁹ *i.e.* a zinc complex of a strapped *N*-alkylporphyrin (Fig. 21). The ELLE of 15 α -amino acids was demonstrated, and it is noteworthy that the extraction of *N*-Cbz-phenylglycinate was performed with a ratio of the host–guest diastereomers of 96:4 in the organic phase.

The unstrapped zinc-porphyrin host face is blocked by the *N*-substituent (\mathbb{R}^2 in Fig. 21). In the strapped cavity, the complexation of *N*-Cbz protected amino acids is presumed to take place. The proposed structure of the *N*-Cbz α -amino acid with the zinc porphyrin involves the complexation of the carboxylate to the zinc



Fig. 21 Strapped *N*-alkylporphyrin zinc host.

ion, which is considered to be the primary interaction. Hydrogen bonding of the benzoyl functionality of the substrate with the amide groups on the strap was confirmed by NMR and IR studies.

Cobalt(III)salen complexes. Excellent results have been reported with a chiral salen-cobalt(III) complex for the enantioselective reactive extraction of *N*-benzyl α -amino acids.¹²⁰ The chiral salen ligands are well-known for their selectivity in asymmetric catalytic processes such as epoxidation, epoxide opening and kinetic resolution.¹²¹ A Co(III)(salen)(OAc) complex is used for reactive extraction of *N*-benzyl protected α -amino acids (Fig. 22). Enantioselective binding is attributed to a bidentate complexation of the protected α -amino acid side group with respect to the salen ligand. The best results imply the extraction of 0.99 equivalents of *N*-Bn-alanine with an enantiomeric excess of 93%. Other amino acids gave ee values in the range of 90–96%.



Fig. 22 Enantioselective complexation of the chiral cobalt(III)salen host with *N*-benzyl α -amino acids.

In the back-extraction procedure, the free substrate was obtained *via* reductive cleavage using 10 eq. sodium dithionite. The formed Co(II) complex can be air-oxidized and re-used without loss of activity. Further studies were performed on the dependence of the protecting group¹²² and the extraction of β^3 -amino acids.¹²³ The reductive back-extraction using sodium dithionite is a major drawback with respect to future application.

Palladium BINAP complexes. Verkuijl *et al.* developed an extraction system based on palladium BINAP complexes. Palladium BINAP complexes are able to extract amino acids enantioselectively with an α_{op} up to 2.8 for tryptophan,⁷⁴ which is the highest reported selectivity for the extraction of tryptophan by metal complexes. The proposed extraction mechanism is according to an interface model, as is shown in Fig. 23.

The use of xyl-BINAP gave even better results, with the α_{op} up to 7.0 for phenylalanine analogues, which is the highest reported selectivity for these class of substrates.¹²⁴ A bidentate binding of the amino acid substrate with the palladium complex was proposed. The extraction of the product of an enzyme catalyzed reaction



Fig. 23 Proposed extraction model for amino acid extraction by palladium BINAP complexes.⁷⁴ © 2009 ACS.

with a chiral palladium complex even facilitated the isolation of enantiopure β -phenylalanine in high yields.¹²⁵

Tartrate assisted extractants

Dialkyl tartrates and boric acid. Abe *et al.* reported the enantioselective extraction of important amino alcohols such as pindolol, propranolol, alprenolol and bucumolol (all β -blockers) using dialkyl L-tartrates in chloroform with boric acid in the aqueous phase¹²⁶ and the use of countercurrent extraction to improve the performance for propanolol.¹²⁷ The optimized operational selectivity of the system was 2.71. Titration of boric acid to a biphasic system with only the tartaric acid present did not show any formation of the dialkyl tartrate boric acid complex. The addition of the amino alcohol substrates resulted in the extraction of the boric acid with the dialkyl tartrate and the amino alcohol substrate. Apparently, in the organic phase the complexes are formed from the dialkyl tartrate boric acid complex and the amino alcohols as bidentate ligands (Fig. 24).



Fig. 24 Proposed structure of the complex formed by boric acid, dialkyl tartrate and amino alcohol substrate.

Viegas *et al.*³⁶ modeled this system based on propanolol/boric acid and di-*n*-dodecyl tartrates in chloroform. The mathematical approach developed involves the use of a hybrid model, with an experimentally determined component, describing the material balance equations of the reactions involved, and a neural network, which allows accounting for the variable non-selective partitioning of propranolol. The modeled intrinsic selectivity was 2.77, indicating that the operational selectivity achieved experimentally by Abe *et al.* (2.71)¹²⁷ was close to the intrinsic optimum of the system.

Di(2-ethylhexyl)phosphoric acid and a tartaric acid derivative. Di(2-ethylhexyl)phosphoric acid (D2EHPA) (Fig. 25) as extractant has been known for over 50 years and has been studied extensively as a racemic mixture of diastereomers in the achiral liquid–liquid extraction of metal cations¹²⁸ and α -amino acids.¹²⁹ A



Fig. 25 Structure of di(2-ethylhexyl)phosphoric acid (D2EHPA).

mechanistic study has been reported by Dai *et al.* for the extraction of L-tryptophan,¹³⁰ which led to the proposal of a proton-transfer reaction occurring in the extraction of the tryptophan.

Luo et al. developed a synergetic system comparable to the dialkyl tartrate-boric acid system as described above,¹³¹ although this system is different, as the tartaric acid component is the free acid, instead of an ester. The D2EHPA is able to extract the tryptophan, albeit without any enantioselectivity. On addition of the tartaric acid, the extraction becomes enantioselective, which makes the system synergetic. The authors propose that the extracting species is a complex formed between D2EHPA and the tartaric acid derivative (O, O'-dibenzoyl-(2R, 3R)-tartaric acid, (-)-DBTA). The complex was employed in the extraction of Trp and the operational selectivity achieved a maximum of $5.3.^{132}$ Ee's up to 57% in the aqueous phase were measured. However, this includes partial diastereomeric salt formation, in other words classic resolution. Luo et al. also published the synergistic extraction using tartrates with Aliquat 336 (trialkyl methyl ammonium chloride, containing mixtures of C8-C10 alkyl chains) with comparable results.133

Diphosphonium salt bearing binaphthyl hosts

A study was reported by Ohki *et al.*¹³⁴ on diphosphonium salts that form a complex with (–)-*O*-dibenzoyltartrate. The structure of this complex is shown in Fig. 26.



Fig. 26 Structure of the diphosphonium salt complex with (-)-O-dibenzoyltartrate.

A reasonable operational selectivity of 1.36 was obtained. Experiments with a monophosphonium salt (one phosphonium salt at the 2- position of the binaphthyl backbone was replaced by a phosphate) showed almost no selectivity, demonstrating the necessity of the bifunctionality of the host.

Alkyl tartrate with β -cyclodextrin. An ELLE system for the separation of mandelic acid (Fig. 28a) was presented by Huang *et al.*, which involves the use of two extractants, with each extractant present in a different phase of the biphasic system.¹³⁵

L-Dipentyl tartrate was used as the extractant confined in the organic phase, while β -cyclodextrin (Fig. 27) was used as the extractant confined in the aqueous phase. The β-cyclodextrin has an enantiopreference which is opposite in the chiral recognition compared to the tartrate. This biphasic recognition increases the selectivity of the ELLE of the substrate mandelic acid up to 2.1. A similar approach was reported for α -cyclohexyl-mandelic acid (Fig. 28b), and naproxen (Fig. 28c).¹³⁶ Selectivities up to 2.49 were reported for these systems with the biphasic extraction system mentioned above. Tang and coworkers have further expanded the scope of substrates to phenylsuccinic acid (selectivity of 2.8),¹³⁷ and the chiral drugs flurbiprofen,138 ibuprofen,139 and zopiclone,140 all with rather low selectivities below 1.5. Much lower but still reasonable selectivities of 1.46-1.85 were observed for the same substrates with a single extractant $Cu(II)(C12Pro)_2$ in the organic phase.90



Fig. 27 β -Cyclodextrin (a) and L-dipentyl tartrate (b), applied as chiral hosts in biphasic recognition.



Fig. 28 Mandelic acid (a), α -cyclohexyl mandelic acid (b) and naproxen (c).

Other chiral extractants

Carbamovlated quinine derivatives. Chiral carbamovlated quinine derivatives have been successfully applied as a chiral stationary phase. Covalently bound to silica gel these compounds have been used in direct enantioseparation by HPLC of chiral acidic compounds employing aqueous mobile phases involving an anionexchange mechanism.¹⁴¹ The selectivity of the enantioselective extraction of racemic Cinchona alkaloids using camphorsulfonic acid as a host is rather low (maximum 1.21). The extractant in its neutral form is soluble in both liquid phases. As a result, the reaction between host and guest may take place in both phases,¹⁴² and this is expected to result in a reduction of the selectivity. Lindner et al. derivatized chiral Cinchona alkaloids as hydrophobic extractants for application in ELLE.¹⁴³ The reactive extraction capabilities of a quinine derivative as a function of the substrate, organic solvent, pH of the water phase and host/guest ratio were examined. Under optimum conditions, an ee > 95% and 70%yield of N-(3,5-dinitrobenzoyl) protected leucine with a single extraction followed by a back-extraction was possible. The system is highly dependent on the 3,5-dinitrobenzoyl (DNB) protecting group on the leucine. On variation of the protecting group, the ee decreases to 20% or less. The π–π interaction between the DNB protecting group and the quinine of the host seems to be vital for enantioselectivity. The ensemble of interactions proposed to be responsible for the chiral discrimination is shown in Fig. 29. The ion-pair formation results in the strongest interaction in the complex. The other interactions shown in Fig. 29 were confirmed by NMR, NOE. and X-ray studies.¹⁴⁴ The high selectivities and versatility towards DNB-substituted α-amino acids make the modified *Cinchona* alkaloids attractive for further research. Various studies regarding the preferred conformation¹⁴⁵⁻¹⁴⁷ and enantioselective pathways^{60,148} for (derivatized) *Cinchona* alkaloids have been reported.



Fig. 29 The interactions proposed to be responsible for the chiral discrimination in extraction of DNB-leucine by a modified *Cinchona* alkaloid extractant.

De Vries *et al.* studied a number of *Cinchona* alkaloids using two continuous centrifugal separators (CCS, see Fig. 30).⁸⁹ The use of continuous equipment was thus demonstrated in a configuration resembling the classical resolving machine of Cram.⁸⁷ Heeres *et al.* have applied a continuous single stage extraction approach using the same type of equipment,⁸² and also showed that the ligand addition mechanism with homogeneous organic phase complexation applies to extractions of DNB substituted α -amino acids using *Cinchona* alkaloid extractants.⁷⁵ It was demonstrated that equilibrium modeling is applicable to describe ELLE in a continuous operation mode.⁸²



Fig. 30 The setup used by De Vries *et al.*⁸⁹ incorporating continuous centrifugal extractors (CCS) to facilitate phase transfer.

Heeres *et al.* have also developed a multistage countercurrent cascade process for the separation of DNB-leucine enantiomers.^{91,92} This was the first time that ELLE was applied in a continuous countercurrent mode, enabling the complete separation of racemates into single enantiomers. A scheme of the setup that was applied is displayed in Fig. 31.



Fig. 31 Fractional six stage countercurrent ELLE process setup with host recycle in back-extraction stage (BE) and feed (F) at stage five as used by Schuur *et al.*⁹² © 2009 ACS.

A single back-extraction stage was proven sufficient for complete recovery of the extracted DNB-(S)-Leu, and the recycling of the host to the extraction section. The host was recycled up to 50 times without loss of enantiomeric excess in the process. With the thus validated multistage equilibrium modeling approach, it was determined that up to 17.7 kg racemate may be separated completely into its enantiomers using 12 units of the bench scale CCS type of equipment. In this process, both enantiomers are obtained with an ee exceeding 99%.^{91,92}

Steroidal guanidinium hosts. The guanidinium functional group has been used in various systems, including the aforementioned de Mendoza host.¹⁰⁰ Davis and co-workers functionalized cholic acid with a guanidinium group at the 3 position and modified the secondary alcohols at the 7 and 12 position to carbamates to obtain a range of chiral hosts (Fig. 32).



Fig. 32 Steroidal guanidinium host.

The phenyl-substituted carbamates (R^1 and R^2 , Fig. 32) gave high enantioselectivities in extraction. These hosts were capable of extracting *N*-acyl protected α -amino acids into the organic phase.^{149,150} The extraction is thought to take place *via* a mechanism involving exchange of the chloride for the carboxylate. The diastereomeric ratio was determined with ¹H-NMR spectroscopy. The ratio of the extracted substrate enantiomers was up to 10:1 (L:D). The most sterically hindered substrate (*N*-Ac-*tert*leucine) gave the lowest selectivity. Enantioselective transport was performed with these type of hosts, using a U-tube and a hollowfiber set-up.¹⁵¹ The host showed multiple turnovers during the transport and the transported substrate present in the receiving phase had an ee of around 60%.

Deoxyguanosine derivatives. The deoxyguanosine derivatives shown in Fig. 33, left, form G-quartet aggregates by self-assembly (Fig. 33, right).¹⁵² In the example published by Spada *et al.*, the R groups are long alkyl chains. Consequently, the outside of the assembly is lipophilic, whereas the inside with the hydrogen bonds is hydrophilic. The intermolecular complexation of these aggregates with chiral guests is most likely to be highly reversible and selectivities up to 3 were reported for the potassium salts of 2,4-DNB- α -amino acids.¹⁵³



Fig. 33 Deoxyguanosine derivative (left) and quartet structure formed by self-assembly (right). $R = p-(n-C_{12}H_{25}O)C_6H_4$.

Sapphyrin–lasalocid host. Another host designed for α -amino acid recognition and membrane transport is the sapphyrin–lasalocid system shown in Fig. 34. Sessler *et al.* designed this host on the premises of non-self-complementary binding sites, which would prevent the host from internal collapse, but could specifically bind the α -amino acid substrates. The protonated sapphyrin can bind to the carboxylate of the α -amino acid, while the lasalocid subunit, known for its transport capabilities of cations through bulk liquid membranes,¹⁵⁴ can complex to the protonated amino group.^{155,156} In U-tube experiments focussing on non-enantioselective transport, a large difference in transport of amino acids was observed.¹⁵⁷ Phenylalanine was transported



Fig. 34 Sapphyrin-lasalocid host.

about 5 times faster compared to tryptophan. In enantioselective transport experiments, the selectivity was low, but the L- α -amino acids were transported faster than the D- α -amino acids. The largest difference observed was for phenylalanine with a 1.4 : 1 (L-Phe : D-Phe) ratio.

TRISPHAT salts. Chiral tris(diimine)ruthenium(II) complexes (Fig. 35, right) are interesting structures because of their special photochemical¹⁵⁸ and biological properties.¹⁵⁹ The complexes can be used for the photo-activated cleavage of specific DNA fragments¹⁵⁹ and in photochemical molecular devices.¹⁶⁰ The enantiopure complexes are ususally obtained *via* classic resolution¹⁶¹ or ion-pair chromatography.¹⁶² Lacour *et al.*¹⁶³ studied the use of chiral TRISPHAT anions (Fig. 35, left) as hosts in ELLE for the separation of the racemic ruthenium complexes¹⁶⁴ and employed them as chiral shift reagents.¹⁶⁵ The chiral lipophilic TRISPHAT anions showed a high enantioselectivity for tris(diimine)ruthenium(II) complexes and values up to 49 were reported.¹⁶³ Data on the reversibility of the complexation reaction and substrate versatility were not provided.



Fig. 35 TRISPHAT anion (left) and tris(diimine)ruthenium(II) complexes (right).

Hydrogen-bond assisted BINOL-aldehyde hosts. Recently, Tang et al.¹⁶⁶ presented two promising hosts containing binaphthyl moieties (Fig. 36) for the ELLE of amino alcohols. The extraction relies on a reversible imine formation of the primary amine of the substrate with the aldehyde of the host, combined with the hydrogen bonding of the alcohol of the substrate with either the urea or the guanidinium of the host. Moderate intrinsic selectivities (3-5) in the complexation with amino-alcohols were obtained with the urea functionalized host (Fig. 36, left).¹⁶⁷ With the guanidinium functionalized host (Fig. 36, right), high intrinsic selectivities for a range of amino alcohols in aqueous/organic biphasic systems with chloroform and benzene as organic phase were achieved. The best results were obtained for 2-amino-1-propanol ($\alpha_{int} = 11$) and 2-amino-1-butanol ($\alpha_{int} =$ 15). Back-extraction of the host was achieved by performing a pH-shift.



Fig. 36 BINOL-aldehyde hosts with a urea (left) and a guanidinium (right) functionality.

The preferred conformations of both diastereomers of the host– guest complexes shown in Fig. 37 provide an explanation for the large difference in selectivity. The steric repulsion between hydrogen and alkyl/aryl around the imine bond of the host and the (S)-amino-alcohol (Fig. 37, left) is larger than that in the case of the (R)-amino-alcohol when the only hydrogen atoms are eclipsed (Fig. 37, right).



Fig. 37 Proposed complexes of the guanidinium BINOL–aldehyde host with (*S*)-amino-alcohols (left) and (*R*)-amino-alcohols (right).

3,3'-Diaryl-BINOL phosphoric acids. 3,3'-Diaryl-BINOL phosphoric acids (Fig. 38, left) were employed as hosts in the enantioselective extraction of primary amines (Fig. 38, right) by Verkuijl *et al.*¹⁶⁸



Fig. 38 Diaryl-BINOL phosphoric acid extractant (left) and primary amine substrate (right).

Operational selectivities as high as 2.2 were achieved in the extraction of phenylglycinol. Titration studies by NMR and FTIR provided evidence for the formation of a 1 : 1 host–guest complex. U-tube experiments proved the reversibility of the system.

Summary for hosts

The hosts discussed in this paper are listed in Table 1.

Conclusions and outlook

The field of enantioselective extraction is undergoing a revival. For decades, satisfying operational selectivities could only be achieved with chiral hosts containing crown ether-like structures. Despite excellent selectivities in several cases the drawback of these systems is the limited scope restricted to primary amines or their ammonium salts. Furthermore, the complex multi-step synthesis restricts access to sufficient amounts of host. With the lanthanide(III) tris(β -diketonate) complexes and the carbamoylated quinine derivatives introduced about ten years ago and the salen-cobalt(III) complexes reported in 2006, it has been shown that reactive extraction does not need to be limited to crown ether systems to achieve good operational selectivities. Furthermore, the experimental verification of the separation of a racemate in a multistage countercurrent cascade demonstrated the proof of principle of the separation of a racemate using a host with intermediate selectivity.

The scalability, recyclability and easy handling of the process makes ELLE in combination with multistage countercurrent cascade technology a fierce rival in the field of new chiral separation technologies. However, there are still major challenges left in the field. Most systems show moderate operational selectivities and a limited substrate scope. For industrial applications, a synthetically acceptably accessible host with a high selectivity towards a wide substrate scope is important to reduce the development time for the separation processes.

The search for new and better host systems is challenging. De Haan *et al.* made an excellent survey on various chiral separator systems used in chromatography and electrophoresis.²⁷ It turns out to be difficult to transfer the existing systems used in these

Table 1 Overview of host	ts
--------------------------	----

Extractant	Substrate	Scope (α_{op}) (-) or [ee](%)	References
Cram dilocular host	α -Amino acid and α -amino acid ester salts	5 examples $(1.5-12)^a$, 18 examples $(1.0-31)^a$	87, 96, 97
BINOL-crown ether	α -Amino acids and ester salts	6 examples $(1.1-19.5)^a$	98
Guanidium-crown ether	α-Amino acids	1 example [up to 30%]	99
Lipophilic crown ethers	Picrate ammonium salts	2 examples (2.2-3.0)	100, 101
Azophenolic crown ethers	Primary amines	6 examples $(1-12)^a$	45-47
Proline derivative Cu(II) complexes	α-Amino acids	5 examples (1.0–4.5)	44, 109–113
Lanthanide(III) tris(β -diketonate) complexes	α-Amino acids and amino alcohols	6 examples (up to 2.2)	114, 117
Metalloporphyrins	N-Cbz α -amino acids	15 examples [50 : 50–96 : 4]	119
Palladium-BINAP complexes	α-Amino acids	22 examples (1.1–7.0)	74, 124
Salen-cobalt(III) complexes	N-Acyl α - and β -amino acids	9 examples [90–96%]	120, 122, 123
Dialkyl tartrates and boric acid	β-Amino alcohols	4 examples (2.20–2.54)	36, 126, 127
D2EHPA and tartaric acid derivative	Amino acids	4 examples (1.36–5.3)	131, 132
Diphosphonium salt	Tartaric acid derivative	1 example (1.36)	134
Alkyl tartrate with β-cyclodextrin	Carboxylic acids, cyclopyrrolone derivate	7 examples (1.20–2.8)	135–140
Carbamoylated quinine Derivatives	N-Protected α -amino acids	Various examples (3–5)	75, 82, 89, 91, 92, 143
Steroidal guanidinium	N-Acyl amino acids	8 examples [1:1–10:1]	149–151
Deoxyguanosine derivatives	DNB-N-protected α -amino acids	6 examples (1.15–3.03)	153
Sapphyrin-lasalocid	α-Amino acids	Low selectivity	157
TRÌSPHAT	Tris(diimine)ruthenium(II) complexes	2 examples [12:1–49:1]	163, 164
3-Aldehyde substitued BINOL	Amino alcohols	7 examples $(1.0-15)^{a}$	166, 167
Diaryl-BINOL phosphoric acid	Primary amines	1.0–2.2	168

^a Intrinsic selectivity.

separation technologies to application in ELLE, mainly because most intrinsic selectivities in these technologies are below 1.2, whereas for ELLE a minimum selectivity of 1.5 is desired to limit the number of extraction stages. Moreover, chromatography usually relies on interactions at the solid–liquid interface. The solid support has a large influence on the mechanism of complexation, which causes the selectivity to be different from the selectivity in a liquid–liquid environment.

New opportunities may come from the field of enantioselective catalysis. Homogeneous asymmetrically catalyzed reactions take place in a fully liquid environment by definition and show unprecedented high enantioselectivities using a wide range of chiral catalysts. These catalysts can be the inspiration for the design of new chiral hosts applied in ELLE. Also, little attention has been paid to systematic variation of functional groups of a specific host. The variation mainly is focussed on the solubility in the organic phase of the host, but seldom to enhance enantiospecific interactions. It is this synthetic approach that deserves more attention.

Another point to stress is the clarification of the mechanism of various systems and the relationship between intrinsic and operational selectivity. Most systems seem to be clear examples of fully reversible systems. Operational selectivity shows dependence on for instance pH, initial concentrations of both substrates and hosts, type of buffer, organic solvents and temperature. It is rarely clear if this dependency between operation selectivity and the various parameters can be adequately explained with either the ligand exchange mechanism or the homogeneous ligand addition mechanism. Both models would imply a strong relation between the intrinsic selectivity and the operational selectivity. This relation is frequently seen in various extraction systems and can open doors towards large scale substrate scanning by examining the intrinsic selectivity with spectroscopic methods and the subsequent optimization of extraction conditions. Little effort has been made towards this goal.

In conclusion, chiral separation by reactive liquid–liquid extraction is an exciting research field, which already has shown proof of principle. Major challenges remain, especially the development of new hosts, which are easily accessible and show a high selectivity for a wide substrate scope, the study of the extraction mechanisms, the examination of the intrinsic selectivities which would enable easy substrate scanning and the application of new systems in a fractional countercurrent multistage cascade to achieve full separation of the substrate enantiomers in a continuous fashion.

Acknowledgements

This work was financially supported by the Netherlands Organization for Scientific Research (NWO-CW) through the Separation Technology program (project nr. 700.54.654) in cooperation with DSM and Schering-Plough.

Notes and references

- 1 D. J. Ager, *Handbook of Chiral Chemicals*, Marcel Dekker, New York, 2005.
- 2 G. M. Ramos Tombo and D. Bellus, *Angew. Chem., Int. Ed. Engl.*, 1991, **30**, 1193–1215.
- 3 R. Noyori, Asymmetric Catalysis in Organic Synthesis, Wiley, New York, 1994.

- 4 R. A. Sheldon, *Chirotechnology*, Marcel Dekker, New York, 1993.
- 5 A. N. Collins, G. N. Sheldrake, and J. Crosby, *Chirality in Industry II*, New York, Wiley and Sons, 1997, p. 411.
- 6 P. K. Ajikumar, K. Tyo, S. Carlsen, O. Mucha, T. H. Phon and G. Stephanopoulos, *Mol. Pharmaceutics*, 2008, **5**, 167–190.
- 7 M. J. Waites, *Industrial Microbiology*, Blackwell Science, Oxford, 2001.
- 8 M. Reschke and K. Schugerl, Chem. Ing. Tech., 1984, 56, 141.
- 9 D. Cascaval, C. Oniscu and A. I. Galaction, *Biochem. Eng. J.*, 2001, 7, 171–176.
- 10 E. N. Jacobsen, A. Pfalz, and H. Yamamoto, *Comprehensive Asymmetric Catalysis*, Springer, Berlin, 1999.
- 11 J. G. de Vries and A. H. M. de Vries, Eur. J. Org. Chem., 2003, 799-811.
- 12 N. M. Maier, P. Franco and W. Lindner, J. Chromatogr., A, 2001, 906, 3–33.
- 13 A. Bruggink, Rational Design in Resolutions, in *Chirality in Industry II*, ed. A. N. Collins, G. N. Sheldrake and J. Crosby, John Wiley & sons Ltd., Chichester, 1997, pp. 81–98.
- 14 F. Faigl, E. Fogassy, M. Nogradi, E. Palovics and J. Schindler, *Tetrahedron: Asymmetry*, 2008, 19, 519–536.
- 15 E. Fogassy, M. Nogradi, D. Kozma, G. Egri, E. Palovics and V. Kiss, Org. Biomol. Chem., 2006, 4, 3011–3030.
- 16 D. Kozma, CRC Handbook of Optical Resolutions Via Diastereomeric Salt Formation, CRC Press LLC, Boca Raton, 2002.
- 17 M. Leeman, G. Brasile, E. Gelens, T. Vries, B. Kaptein and R. Kellogg, *Angew. Chem.*, *Int. Ed.*, 2008, **47**, 1287–1290.
- 18 S. Houlton, Manuf. Chem., 2001, 72, 16-17.
- 19 Y. Fujima, M. Ikunaka, T. Inoue and J. Matsumoto, Org. Process Res. Dev., 2006, 10, 905–913.
- 20 S. Ahuja, *Chiral Separations: Application and Technology*, ACS, Washington, 1997.
- 21 G. Subramanian, Chiral Separation Techniques: A Practical Approach, Wiley-VCH, Weinheim, 2001.
- 22 F. Toda, *Enantiomer Separation: Fundamentals and Practical Methods*, Kluwer Academic Publishers, Dordrecht, 2004.
- 23 G. Guebitz and M. G. Schmid, *Chiral Separations*, Humana Press, Totowa (NJ), 2004.
- 24 K. W. Busch and M. A. Busch, *Chiral Analysis*, Elsevier, Amsterdam, 2004.
- 25 V. A. Davankov, J. Chromatogr., A, 1994, 666, 55-76.
- 26 T. Jira, A. Bunke, M. G. Schmid and G. Gubitz, J. Chromatogr., A, 1997, **761**, 269–275.
- 27 M. Steensma, N. J. M. Kuipers, A. B. de Haan and G. Kwant, *Chirality*, 2006, 18, 314–328.
- 28 G. B. Cox, *Preparative Enantioselective Chromatography*, Blackwell Publishing, Oxford, 2005.
- 29 E. Gavioli, N. M. Maier, C. Minguillon and W. Lindner, *Anal. Chem.*, 2004, **76**, 5837–5848.
- 30 E. Francotte, T. Leutert, L. La Vecchia, F. Ossola, P. Richert and A. Schmidt, *Chirality*, 2002, 14, 313–317.
- 31 G. Zenoni, F. Quattrini, M. Mazzotti, C. Fuganti and M. Morbidelli, *Flavour Fragrance J.*, 2002, 17, 195–202.
- 32 C. A. M. Afonso and J. G. Crespo, Angew. Chem., Int. Ed., 2004, 43, 5293–5295.
- 33 J. T. F. Keurentjes, L. J. W. M. Nabuurs and E. A. Vegter, J. Membr. Sci., 1996, 113, 351–360.
- 34 A. Maximini, H. Chmiel, H. Holdik and N. W. Maier, J. Membr. Sci., 2006, 276, 221–231.
- 35 R. Xie, L. Y. Chu and J. G. Deng, *Chem. Soc. Rev.*, 2008, **37**, 1243–1263.
- 36 R. M. C. Viegas, C. A. M. Afonso, J. G. Crespo and I. M. Coelhoso, Sep. Purif. Technol., 2007, 53, 224–234.
- 37 A. B. De Haan and B. Simandi, Extraction Technology for the Separation of Optical Isomers, in *Ion Exchange and Solvent Extraction*, ed. Y. Marcus, M. M. Sharma and J. A. Marinsky, Marcel Dekker, Inc, New York, 2001, pp. 255–294.
- 38 L. B. Dashkevic, Tr. Leningr. Khim.-Farm. Inst., 1959, 6, 29.
- 39 K. Bauer, H. Falk and K. Schlogl, Monatsh. Chem., 1968, 99, 2186– 2194.
- 40 N. S. Bowman, G. T. Mccloud and G. K. Schweitz, J. Am. Chem. Soc., 1968, 90, 3848–3852.
- 41 S. J. Romano, K. H. Wells, H. L. Rothbart and W. Rieman, *Talanta*, 1969, 16, 581–590.

- 42 G. K. Schweitz, I. R. Supernaw and N. S. Bowman, J. Inorg. Nucl. Chem., 1968, 30, 1885–1890.
- 43 J. Koska and C. A. Haynes, Chem. Eng. Sci., 2001, 56, 5853-5864.
- 44 P. J. Pickering and J. B. Chaudhuri, *Chem. Eng. Sci.*, 1997, **52**, 377–386.
- 45 M. Steensma, N. J. Kuipers, A. B. de Haan and G. Kwant, J. Chem. Technol. Biotechnol., 2006, 81, 588–597.
- 46 M. Steensma, N. J. M. Kuipers, A. B. de Haan and G. Kwant, *Chem. Eng. Sci.*, 2007, 62, 1395–1407.
- 47 M. Steensma, N. J. M. Kuipers, A. B. de Haan and G. Kwant, *Chem. Eng. Proc.*, 2007, 46, 996–1005.
- 48 E. Eliel, S. Wilen and L. Mander, Miscellaneous Separation Methods, in *Stereochemistry of organic compounds*, Wiley, New York, NY, 1994, pp. 416–424.
- 49 J. C. Godfrey and M. J. Slater, *Liquid–Liquid Extraction Equipment*, John Wiley & Sons, New York, 1994.
- 50 D. J. Cram and J. M. Cram, *Container Molecules and Their Guests*, Royal Society of Chemistry, Cambridge, 1994.
- 51 F. Toda, K. Tanaka, C. W. Leung, A. Meetsma and B. L. Feringa, J. Chem. Soc., Chem. Commun., 1994, 2371–2372.
- 52 K. Yoshizawa, S. Toyota and F. Toda, *Tetrahedron*, 2004, **60**, 7767–7774.
- 53 F. Toda, K. Yoshizawa, S. Hyoda, S. Toyota, S. Chatziefthimiou and I. M. Mavridis, Org. Biomol. Chem., 2004, 2, 449–451.
- 54 J. Liao, X. X. Sun, X. Cui, K. B. Yu, J. Zhu and J. G. Deng, *Chem.-Eur. J.*, 2003, 9, 2611–2615.
- 55 G. Kaupp, Angew. Chem., Int. Ed. Engl., 1994, 33, 728-729.
- 56 W. H. Pirkle and T. C. Pochapsky, Chem. Rev., 1989, 89, 347-362.
- 57 T. C. Lo, M. H. I. Baird, and C. Hanson, *Handbook of Solvent Extraction*, Krieger Publishing Company, Malabar, USA, 1991.
- 58 J. D. Thornton, *Science and Practice of Liquid–Liquid Extraction*, Clarendon, Oxford, 1992.
- 59 R. E. Treybal, *Liquid Extraction*, 2nd Ed., McGraw-Hill, New York, 1963.
- 60 N. M. Maier, S. Schefzick, G. M. Lombardo, M. Feliz, K. Rissanen, W. Lindner and K. B. Lipkowitz, *J. Am. Chem. Soc.*, 2002, **124**, 8611– 8629.
- 61 V. A. Davankov, Chirality, 1997, 9, 99–102.
- 62 V. Sundaresan and R. Abrol, Chirality, 2005, 17, S30-S39.
- 63 E. L. Eliel and S. H. Wilen, Stereochemistry of Organic Compounds, Wiley-Interscience, New York, 1994.
- 64 J. M. Lehn, Supramolecular Chemistry: Concepts and Perspectives, Weinheim, VCH, 1995.
- 65 J. H. Hartley, T. D. James and C. J. Ward, J. Chem. Soc., Perkin Trans. 1, 2000, 3155–3184.
- 66 P. D. Beer and P. A. Gale, Angew. Chem., Int. Ed., 2001, 40, 486–516.
- 67 I. Stibor and P. Zlatuskova, Anion Sensing, 2005, 255, 31-63.
- 68 C. Seel, A. Galan and J. deMendoza, Supramolecular Chemistry II -Host Design and Molecular Recognition, 1995, 175, 101–132.
- 69 M. W. Peczuh and A. D. Hamilton, *Chem. Rev.*, 2000, **100**, 2479–2493.
- 70 A. P. Davis and R. S. Wareham, *Angew. Chem., Int. Ed.*, 1999, **38**, 2978–2996.
- 71 R. J. Fitzmaurice, G. M. Kyne, D. Douheret and J. D. Kilburn, J. Chem. Soc., Perkin Trans. 1, 2002, 841–864.
- 72 T. H. Webb and C. S. Wilcox, Chem. Soc. Rev., 1993, 22, 383–395.
- 73 X. X. Zhang, J. S. Bradshaw and R. M. Izatt, Chem. Rev., 1997, 97,
- 3313–3361.
 74 B. J. V. Verkuijl, A. J. Minnaard, J. G. de Vries and B. L. Feringa, J. Org. Chem., 2009, 74, 6526–6533.
- 75 B. Schuur, J. G. M. Winkelman and H. J. Heeres, *Ind. Eng. Chem. Res.*, 2008, **47**, 10027–10033.
- 76 H. J. Bart, Reactive Extraction, Springer Verlag, Berlin, 2001.
- 77 M. Cox, Liquid–Liquid Extraction in Hydrometallurgy, in *Science and Practice of Liquid–Liquid Extraction*, ed. J. D. Thornton, Clarendon, Oxford, 1992, pp. 1–101.
- 78 C. J. King, Chemtech, 1992, 22, 285-291.
- 79 K. L. Wasewar, A. A. Yawalkar, J. A. Moulijn and V. G. Pangarkar, *Ind. Eng. Chem. Res.*, 2004, **43**, 5969–5982.
- 80 G. F. Cegla, W. W. Meinke and K. F. Mattil, J. Food Sci., 1977, 42, 816–820.
- 81 M. Tanaka, K. Koyama and J. Shibata, Ind. Eng. Chem. Res., 1997, 36, 4353–4357.
- 82 B. Schuur, J. Floure, A. J. Hallett, J. G. M. Winkelman, J. G. de Vries and H. J. Heeres, Org. Process Res. Dev., 2008, 12, 950–955.

- 83 G. S. Laddha and T. E. Degaleesan, *Transport Phenomena in Liquid Extraction*, Tata McGraw-Hill, New Delhi, 1978.
- 84 M. Hafez, Centrifugal Extractors, in *Handbook of Solvent Extraction*, ed. T. C. Lo, M. H. C. Baird, and C. Hanson, Krieger Publishing Company, Malabar, 1991, pp. 459–474.
- 85 L. S. Ettre, J. Chromatogr. Sci., 1999, 37, 2A-8A.
- 86 G. J. Hanna and R. D. Noble, Chem. Rev., 1985, 85, 583-598.
- 87 M. Newcomb, J. L. Toner, R. C. Helgeson and D. J. Cram, J. Am. Chem. Soc., 1979, 101, 4941–4947.
- 88 P. Breccia, M. Van Gool, R. Perez-Fernandez, S. Martin-Santamaria, F. Gago, P. Prados and J. C. de Mendoza, J. Am. Chem. Soc., 2003, 125, 8270–8284.
- 89 A. J. Hallett, G. J. Kwant and J. G. de Vries, *Chem.–Eur. J.*, 2009, **15**, 2111–2120.
- 90 H. Nishizawa, K. Tahara, A. Hayashida and Y. Abe, *Anal. Sci.*, 1993, 9, 611–615.
- 91 B. Schuur, J. G. M. Winkelman, J. G. de Vries and H. J. Heeres, *Chem. Eng. Sci.*, 2010, **65**, 4682–4690.
- 92 B. Schuur, A. J. Hallett, J. G. M. Winkelman, J. G. de Vries and H. J. Heeres, Org. Process Res. Dev., 2009, 13, 911–914.
- 93 C. J. Pedersen, J. Am. Chem. Soc., 1967, 89, 7017-7019.
- 94 C. J. Pedersen, J. Am. Chem. Soc., 1967, 89, 2495-2497.
- 95 J. M. Lehn and A. Moradpour, *Helv. Chim. Acta*, 1978, **61**, 2407–2418.
- 96 E. P. Kyba, J. M. Timko, L. J. Kaplan, F. Dejong, G. W. Gokel and D. J. Cram, J. Am. Chem. Soc., 1978, 100, 4555–4568.
- 97 S. C. Peacock, L. A. Domeier, F. C. A. Gaeta, R. C. Helgeson, J. M. Timko and D. J. Cram, J. Am. Chem. Soc., 1978, 100, 8190–8202.
- 98 D. S. Lingenfelter, R. C. Helgeson and D. J. Cram, J. Org. Chem., 1981, 46, 393–406.
- 99 A. Galan, D. Andreu, A. Echavarren, P. Prados and J. De Mendoza, J. Am. Chem. Soc., 1992, 114, 1511–1512.
- 100 M. Colera, A. M. Costero, P. Gavina and S. Gil, *Tetrahedron:* Asymmetry, 2005, 16, 2673–2679.
- 101 A. Y. Nazarenko, P. Huszthy, J. S. Bradshaw, J. D. Lamb and R. M. Izatt, J. Inclusion Phenom. Mol. Recognit. Chem., 1994, 20, 13–22.
- 102 S. S. Moore, T. L. Tarnowski, M. Newcomb and D. J. Cram, J. Am. Chem. Soc., 1977, 99, 6398–6405.
- 103 K. Naemura, K. Nishioka, K. Ogasahara, Y. Nishikawa, K. Hirose and Y. Tobe, *Tetrahedron: Asymmetry*, 1998, 9, 563–574.
- 104 K. Naemura, K. Matsunaga, J. Fuji, K. Ogasahara, Y. Nishikawa, K. Hirose and Y. Tobe, *Anal. Sci.*, 1998, 14, 175–182.
- 105 P. E. Hare and E. Gil-Av, Science, 1979, 204, 1226-1228.
- 106 G. Gübitz, W. Jellenz and W. Santi, J. Liq. Chromatogr. Relat. Technol., 1981, 4, 701–712.
- 107 V. A. Davankov and S. V. Rogozhin, J. Chromatogr., A, 1971, 60, 280–283.
- 108 G. Gubitz, W. Jellenz and W. Santi, J. Chromatogr., A, 1981, 203, 377–384.
- 109 T. Takeuchi, R. Horikawa and T. Tanimura, Anal. Chem., 1984, 56, 1152–1155.
- 110 P. J. Pickering and J. B. Chaudhuri, Chirality, 1997, 9, 261-267.
- 111 P. J. Pickering and J. B. Chaudhuri, J. Membr. Sci., 1997, 127, 115–130.
- 112 H. B. Ding, P. W. Carr and E. L. Cussler, *AIChE J.*, 1992, **38**, 1493–1498.
- 113 P. Dimitrova and H. J. Bart, Chem. Eng. Technol., 2009, 32, 1527-1534.
- 114 H. Tsukube, J. Uenishi, T. Kanatani, H. Itoh and O. Yonemitsu, *Chem. Commun.*, 1996, 477–478.
- 115 G. R. Sullivan, D. Ciavarel and H. S. Mosher, J. Org. Chem., 1974, 39, 2411–2412.
- 116 C. Kremer, J. Torres, S. Dominguez and A. Mederos, *Coord. Chem. Rev.*, 2005, 249, 567–590.
- 117 H. Tsukube, S. Shinoda, J. Uenishi, T. Kanatani, H. Itoh, M. Shiode, T. Iwachido and O. Yonemitsu, *Inorg. Chem.*, 1998, **37**, 1585–1591.
- 118 R. Rinaldi, Prog. Nucl. Magn. Reson. Spectrosc., 1982, 15, 291– 352.
- 119 K. Konishi, K. Yahara, H. Toshishige, T. Aida and S. Inoue, J. Am. Chem. Soc., 1994, 116, 1337–1344.
- 120 T. B. Reeve, J. P. Cros, C. Gennari, U. Piarulli and J. G. de Vries, Angew. Chem., Int. Ed., 2006, 45, 2449–2453.
- 121 J. R. Larrow and E. N. Jacobsen, *Top. Organomet. Chem.*, 2004, 7, 123–152.
- 122 P. Dzygiel, T. B. Reeve, U. Piarulli, M. Krupicka, I. Tvaroska and C. Gennari, *Eur. J. Org. Chem.*, 2008, 1253–1264.
- 123 P. Dzygiel, C. Monti, U. Piarulli and C. Gennari, Org. Biomol. Chem., 2007, 5, 3464–3471.

- 124 B. J. V. Verkuijl, B. Schuur, A. J. Minnaard, J. G. de Vries and B. L. Feringa, Org. Biomol. Chem., 2010, 8, 3045–3054.
- 125 B. J. V. Verkuijl, W. Szymanski, B. Wu, A. J. Minnaard, D. B. Janssen, J. G. de Vries and B. L. Feringa, *Chem. Commun.*, 2010, 46, 901–903.
- 126 Y. Abe, T. Shoji, M. Kobayashi, W. Qing, N. Asai and H. Nishizawa, *Chem. Pharm. Bull.*, 1995, 43, 262–265.
- 127 Y. Abe, T. Shoji, S. Fukui, M. Sasamoto and H. Nishizawa, *Chem. Pharm. Bull.*, 1996, 44, 1521–1524.
- 128 I. Van de Voorde, L. Pinoy, E. Courtijn and F. Verpoort, *Solvent Extr. Ion Exch.*, 2006, 24, 893–914.
- 129 M. Teramoto, T. Yamashiro, A. Inoue, A. Yamamoto, H. Matsuyama and Y. Miyake, J. Membr. Sci., 1991, 58, 11–32.
- 130 Y. S. Liu, Y. Y. Dai and J. D. Wang, Sep. Sci. Technol., 2000, 35, 1439–1454.
- 131 B. Tan, G. S. Luo, X. Qi and J. D. Wang, Sep. Purif. Technol., 2006, 49, 186–191.
- 132 B. Tan, G. S. Luo and H. D. Wang, *Tetrahedron: Asymmetry*, 2006, 17, 883–891.
- 133 B. Tan, G. S. Luo and J. D. Wang, Sep. Purif. Technol., 2007, 53, 330–336.
- 134 A. Ohki, R. Miyashita, K. Naka and S. Maeda, Bull. Chem. Soc. Jpn., 1991, 64, 2714–2719.
- 135 F. P. Jiao, X. Q. Chen, W. G. Hu, F. R. Ning and K. L. Huang, *Chem. Pap.*, 2007, **61**, 326–328.
- 136 K. Tang, Y. Chen, K. Huang and J. Liu, *Tetrahedron: Asymmetry*, 2007, 18, 2399–2408.
- 137 K. W. Tang, J. M. Yi, Y. B. Liu, X. Y. Jiang and Y. Pan, *Chem. Eng. Sci.*, 2009, 64, 4081–4088.
- 138 K. W. Tang, L. T. Song, Y. B. Liu, Y. Pan and X. Y. Jiang, *Chem. Eng. J.*, 2010, **158**, 411–417.
- 139 K. W. Tang, L. T. Song, Y. Pan, X. Y. Jiang and J. B. Miao, *Chin. J. Chem.*, 2010, **28**, 119–124.
- 140 K. W. Tang, Y. Y. Chen and J. J. Liu, Sep. Purif. Technol., 2008, 62, 681–686.
- 141 M. Lammerhofer and W. Lindner, J. Chromatogr., A, 1996, 741, 33–48.
- 142 R. D. Mhaskar and M. M. Sharma, Chem. Eng. Sci., 1975, 30, 811– 818.
- 143 K. H. Kellner, A. Blasch, H. Chmiel, M. Lammerhofer and W. Lindner, *Chirality*, 1997, 9, 268–273.
- 144 C. Czerwenka, M. M. Zhang, H. Kahlig, N. M. Maier, K. B. Lipkowitz and W. Lindner, J. Org. Chem., 2003, 68, 8315–8327.
- 145 H. Caner, P. U. Biedermann and I. Agranat, *Chirality*, 2003, 15, 637–645.

- 146 G. D. H. Dijkstra, R. M. Kellogg and H. Wynberg, J. Org. Chem., 1990, 55, 6121–6131.
- 147 G. D. H. Dijkstra, R. M. Kellogg, H. Wynberg, J. S. Svendsen, I. Marko and K. B. Sharpless, *J. Am. Chem. Soc.*, 1989, **111**, 8069–8076.
- 148 G. Uccello-Barretta, F. Balzano, C. Quintavalli and P. Salvadori, J. Org. Chem., 2000, 65, 3596–3602.
- 149 L. J. Lawless, A. G. Blackburn, A. J. Ayling, M. N. Perez-Payan and A. P. Davis, J. Chem. Soc., Perkin Trans. 1, 2001, 1329–1341.
- 150 A. P. Davis and L. J. Lawless, Chem. Commun., 1999, 9-10.
- 151 B. Baragana, A. G. Blackburn, P. Breccia, A. P. Davis, J. De Mendoza, J. M. Padron-Carrillo, P. Prados, J. Riedner and J. G. de Vries, *Chem.– Eur. J.*, 2002, 8, 2931–2936.
- 152 J. T. Davis, Angew. Chem., Int. Ed., 2004, 43, 668-698.
- 153 V. Andrisano, G. Gottarelli, S. Masiero, E. H. Heijne, S. Pieraccini and G. P. Spada, *Angew. Chem., Int. Ed.*, 1999, **38**, 2386–2388.
- 154 I. Spasojevic and A. L. Crumbliss, J. Chem. Soc., Dalton Trans., 1998, 4021–4027.
- 155 J. L. Sessler and A. Andrievsky, Chem. Commun., 1996, 1119-1120.
- 156 A. Andrievsky and J. L. Sessler, *Abstr. Pap. ACS*, 1996, **211**, 76–77.
- 157 J. L. Sessler and A. Andrievsky, Chem.-Eur. J., 1998, 4, 159-167.
- 158 A. Vonzelewsky, P. Belser, P. Hayoz, R. Dux, X. Hua, A. Suckling and H. Stoecklievans, *Coord. Chem. Rev.*, 1994, 132, 75–85.
- 159 J. K. Barton, Science, 1986, 233, 727-734.
- 160 V. Balzani, A. Credi, and M. Venturi, *Molecular Devices and Machines:* Concepts and Perspectives for the Nanoworld, Wiley-VCH, Weinheim, 2008.
- 161 R. D. Gillard and R. E. E. Hill, J. Chem. Soc., Dalton Trans., 1974, 1217–1236.
- 162 J. Lacour, S. Torche-Haldimann, J. J. Jodry, C. Ginglinger and F. Favarger, *Chem. Commun.*, 1998, 1733–1734.
- 163 J. Lacour, C. Goujon-Ginglinger, S. Torche-Haldimann and J. J. Jodry, Angew. Chem. Int. Ed., 2000, 39, 3695–3697.
- 164 J. J. Jodry and J. Lacour, Chem.-Eur. J., 2000, 6, 4297-4304.
- 165 H. Ratni, J. J. Jodry, J. Lacour and E. P. Kundig, Organometallics, 2000, 19, 3997–3999.
- 166 L. Tang, S. Choi, R. Nandhakumar, H. Park, H. Chung, J. Chin and K. M. Kim, J. Org. Chem., 2008, 73, 5996–5999.
- 167 K. M. Kim, H. Park, H. J. Kim, J. Chin and W. Nam, Org. Lett., 2005, 7, 3525–3527.
- 168 B. J. V. Verkuijl, J. G. de Vries and B. L. Feringa, *Chirality*, 2010, DOI: 10.1002/chir.20834.